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ЕЖЕМЕСЯЧНЫЙ НАУЧНЫЙ ЖУРНАЛ

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**ЕЖЕМЕСЯЧНЫЙ НАУЧНЫЙ ЖУРНАЛ
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Контактный адрес: Грузия, 0177, Тбилиси, ул. Асатиани 7, IV этаж, комната 408
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Fax: +995(32) 253 70 58, e-mail: ninomikaber@geomednews.com; nikopir@geomednews.com

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GMN Editorial Board

Phone: 995 (32) 254-24-91

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995 (32) 253-70-58

Tbilisi, Georgia 0177

Fax: 995 (32) 253-70-58

CONTACT ADDRESS IN NEW YORK

NINITEX INTERNATIONAL, INC.

3 PINE DRIVE SOUTH

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PERIPHERAL BLOOD BIOMARKERS IN PATIENTS WITH REFRACTORY IMMUNE THROMBOCYTOPENIA

¹Metreveli S., ³Kvachadze I., ^{1,2}Kikodze N., ¹Chikovani T., ¹Janikashvili N.

*Tbilisi State Medical University, ¹Department of Immunology;
²Institute of Medical Biotechnology; ³Department of Physiology, Georgia*

Immune thrombocytopenia (ITP, also called idiopathic thrombocytopenic purpura) is an acquired thrombocytopenia caused by autoantibodies against platelet antigens [1]. It is one of the most common causes of thrombocytopenia in otherwise asymptomatic adults. The pathogenesis of ITP is incompletely understood.

Therapy for immune thrombocytopenia (ITP) differs for different patients: some do not require any treatment; others have a spontaneous remission or respond to first-line therapy with glucocorticoids; and others continue to have severe thrombocytopenia necessitating additional therapy [2]. Second-line therapy is generally reserved for patients with thrombocytopenia that are refractory to first-line treatment and is associated with significant bleeding symptoms (e.g., mucosal purpura, more serious bleeding) or for severe, persistent or recurrent thrombocytopenia (e.g., platelet count <20,000/microL) following glucocorticoid-based treatments [3-6]. The group of patients we have investigated in this report was refractory for the first line treatment and had splenectomy as a second-line therapy.

Inflammation is known to have a significant role in the course of many benign and malignant diseases. Neutrophil-lymphocyte ratio (NLR) and Platelet-to-lymphocyte ratio (PLR) have been used frequently as a marker of systemic inflammation in recent years. Liping Wang et al. demonstrated that NLR was significantly higher in systemic lupus erythematosus (SLE) patients than in healthy controls (SMD=1.43; 95% CI, 0.98–1.88) [7]. PLR and NLR are also frequently used in clinical studies for Rheumatoid Arthritis (RA) [8-10]. Most of the investigations of inflammatory factors with ITP patients are associated with the first line treatment. It has been studied the response to corticosteroid therapy associated with NLR in ITP [11], Platelet-to-lymphocyte ratio (PLR) has also been investigated in different studies, risk of recurrence in ITP patients with first-line treatment, correlation between PLR and Glucocorticoid resistance in newly identified ITP [12,13], but evidence regarding association between inflammatory factors in patients with ITP who are refractory to first line treatment is limited.

Herein, we have investigated the altered NLR, PLR, plate-

let to monocyte ratio (PMR), derived NLR (dNLR) and systemic immune-inflammation index (SII) values in ITP patients who had splenectomy as a second line treatment and compare it with age-matched control group and their correlation with platelet count.

Material and methods. This study was carried out in accordance with the principles of the 1975 Declaration of Helsinki and its later amendments or comparable ethical standards, and was approved by Tbilisi State Medical University Biomedical Research Ethics Committee. Formal consent was not required for this retrospective study, while all data were kept confidential.

Systemic inflammatory factors NLR, PLR, PMR, dNLR, and SII were conveyed and analyzed from 6 ITP patients who were refractory to first line treatment and had splenectomy as a second line therapy, and 28 age-matched controls (the patients who had other hematologic disease, cancers, acute or chronic infections, liver or kidney disease, were excluded from the study).

The counts for white blood cells, neutrophil, lymphocyte, platelet and monocyte were taken from patient's CBC analyses extracted from medical records. NLR, PLR, PMR were calculated as the absolute count of neutrophils, platelets, monocytes respectively, divided by the absolute lymphocyte count. The dNLR was calculated using a formula: dNLR = neutrophil/(total white cell - neutrophil). The SII was defined as follows: SII = monocyte X Platelet/lymphocyte.

Statistical analysis was performed using Graph Pad and SPSS software. Correlations between the variables were determined by Spearman's correlation coefficient. The area under the curve (AUC), sensitivity, specificity, and cut-off values were compared using the receiver operating characteristic (ROC) curve. A P<0.05 was considered as statistically significant. Categorical variables were shown as percentages and continuous variables were presented as mean ± standard deviation (SD).

Result and discussion. Patient characteristics and laboratory parameters are summarized in Table 1. The median age of the patients at the time of diagnosis was 49 years (range, 20-76), the genders was equal. None of the ITP patient had response on first line treatment; in all cases were done splenectomy as a second line therapy.

Table 1. Demographic characteristics and laboratory parameters of patients with refractory ITP and age-matched control

Parameters	Patients Group (n = 6)	Control Group (n = 28)	p Value
Age	49.00 ± 9.38	48.3±15.7	NS
Gender: F/M (%)	50/50%	57.1/42.9%	NS
Total white blood cells (×10 ⁹ /L)	8.39 ± 1.968	6.36 ± 1.15	=0.0294
Absolute neutrophil count (×10 ⁹ /L)	5.94 ± 1.65	3.65 ± 0.9	=0.0037
Absolute lymphocyte count (×10 ⁹ /L)	2.53 ± 0.65	2.05 ± 0.4	NS
Absolute monocyte count (×10 ⁹ /L)	0.58 ± 0.19	0.5 ± 0.1	NS
Total platelets (×10 ⁹ /L)	52.33 ± 21.32	248.9 ± 43.9	<0.0001
Hemoglobin	12.47 ± 0.77	13.6 ± 0.97	=0.440

Table 2. Comparison of systemic inflammatory factors in ITP patients and age-matched control groups

Parameters	ITP n=6	Control n=28	P value
NLR	3.763 ± 0.6894	1.849 ± 0.1112	<0.0001
PLR	25.14 ± 5.007	125.5 ± 5.713	<0.0001
PMR	76.60 ± 15.18	511.2 ± 22.96	<0.0001
dNLR	2.750 ± 0.5241	1.366 ± 0.06943	<0.0001
SII	25.13 ± 8.754	63.30 ± 3.985	0.0003

NLR - neutrophil lymphocyte ratio, PLR - platelet lymphocyte ratio, PMR - platelet monocyte ratio,
dNLR - Derived neutrophil to lymphocyte ratio, The SII - Systemic Immune-Inflammation Index

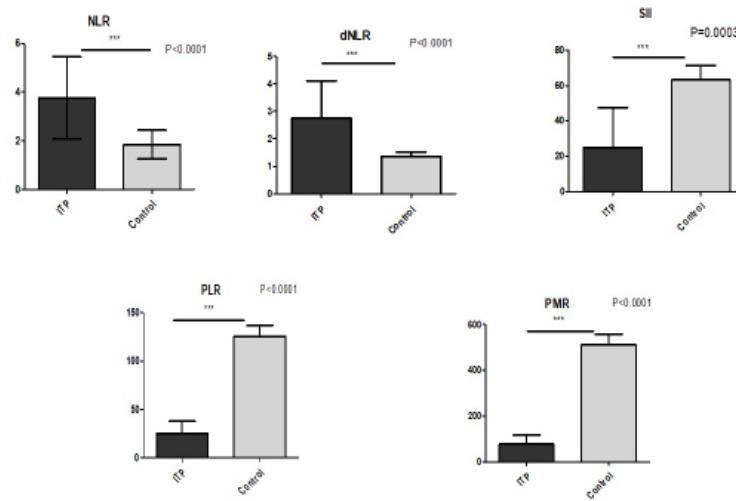


Fig. 1. NLR, PLR, PMR, dNLR, and SII in refractory ITP Patients compared with age-matched control

Table 3. The results of the receiver operating characteristic (ROC) curve analysis of refractory ITP patients compared with age-matched control

	AUC	SE	95% CI	Cut-off	Sensitivity (%)	Specificity (%)	p-value
NLR	0.875	0.104	0.671–1.000	2.64	83.3	99.92	=0.04
PLR	1.000	0.067	1.000–1.000	59.25	100	100	=0.05
PMR	1.000	0.000	1.000–1.000	211.25	100	100	<0.001
dNLR	0.869	0.115	0.644–1.000	1.91	83.3	99.96	<0.001
SII	0.899	0.059	0.782–1.000	47.45	83.3	99.78	=0.002

We explored higher NLR, dNLR in ITP patients than the control group (Fig. 1). The NLR in age-matched control and ITP were 1.849 ± 0.1112 and 3.763 ± 0.6894 ($p<0.0001$); dNLR were 1.366 ± 0.06943 and 2.750 ± 0.5241 ($p<0.0001$); SII were diminished in ITP patients compared to healthy group 25.13 ± 8.754 and 63.30 ± 3.985 ($p=0.0003$), as shown in Table 2, Figure 1. PLR and PMR were significantly low in ITP Patients compared to age-matched control. PLR was 25.14 ± 5.007 and 125.5 ± 5.713 in ITP and control group respectively ($p<0.0001$). PMR was also significantly low in ITP patients compared to control group 76.60 ± 15.18 and 511.2 ± 22.96 ($p<0.0001$).

ROC – receiver-operating characteristic; CI – confidence interval; AUC – area under the curve; SE – standard error; NLR – neutrophil/lymphocyte ratio; PLR – platelet/lymphocyte ratio; PMR: platelet monocytosis ratio; dNLR derived neutrophil-to-lymphocyte ratio; SII: Systemic Immune-Inflammation Index

The results of the receiver operating characteristic (ROC) curve analysis of studied parameters are presented in Table 3. All have an area under the curve of more than 0.5. PLR and

PMR had the highest AUC scores of 1.000 with a 95% CI of 1.000–1.000, followed by SII (0.899), NLR (0.875) and dNLR (0.869).

The Study demonstrates that PLT was negatively correlated with NLR and dNLR $r=-0.605$, P value is below 0.01 level. There was positive correlation with SII $r=0.799$; PLR $r=0.863$; PMR $r=0.40$. P value is also below 0.01 levels (Table 4).

The pathogenesis of ITP is incompletely understood. Reduced platelet lifespan is due to antibody-mediated destruction produced by the patient's B cells, most often directed against platelet membrane glycoproteins such as GPIIb/IIIa; however, other mechanisms are likely important, including autoreactive cytotoxic T cells, as well as humoral and cellular autoimmunity directed at megakaryocytes, causing impaired platelet production. [1][14][15]. Antibody production in ITP appears to be driven by CD4-positive helper T cells reacting to platelet surface glycoproteins, possibly involving CD40:CD40L co-stimulation [16]. Splenic macrophages appear to be the major antigen-presenting cells.

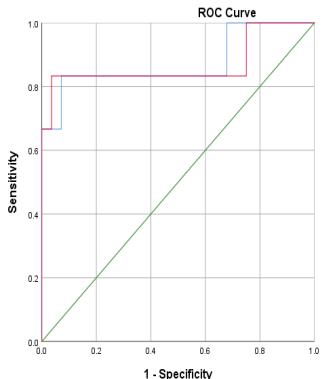


Fig. 2. ROC curves for peripheral blood biomarkers, NLR, dNLR

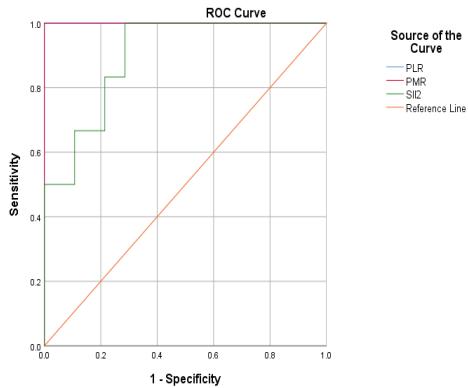


Fig. 3. ROC curves for PLR, PMR and SII

Table 4. Correlation coefficients of inflammatory biomarkers: NLR, PLR, PMR, dNLR, and SII with PLT in refractory ITP

	NLR	dNLR	PLR	SII	PMR	NLR
PLT	-0.605*	-0.605*	0.863*	0.799*	0.840*	-0.605*
P value	0.000	0.000	0.000	0.000	0.000	0.000

PLT- platelets total; NLR – neutrophil/lymphocyte ratio; PLR – platelet/lymphocyte ratio; PMR: platelet monocyt ratio; dNLR – derived neutrophil-to-lymphocyte ratio; SII: Systemic Immune-Inflammation Index.

* - correlation is significant at the 0.01 level

Inflammatory factors have been studied in many autoimmune diseases. Lixiu Li et al demonstrates NLR in SLE patients and compares it with the healthy controls. NLR in SLE patients were significantly elevated and had high diagnostic value (sensitivity, 0.574; specificity, 0.926; 95% CI, 0.668–0.845; P<0.001). [17]

Zunni Zhang et al observed Behcet Disease diagnostic values of NLR, HB and combined NLR with HB and found the value particularly high when combine NLR with HB.[18]

Go Eun Yang et al. showed that in chronic ITP patients NLR level is low at diagnosis (1.23 ± 1.38 vs. 1.54 ± 1.15 , P=0.42) compared after recovery when NLR is significantly high (1.58 ± 1.71 vs. 1.31 ± 0.76 , P=0.21).[8]

In a study conducted by Jun Song et al, they found that PLR may be a useful parameter to consider the risk of recurrence in ITP patients receiving first-line therapy. [13]

As there are limited data of inflammation markers in refractory ITP, in our study we evaluated the inflammation factors NLR, PLR, and MLR, dNLR, and SII in ITP patients who were refractory to first line therapy and had a splenectomy as a second line treatment and compared the results with aged matched control group.

In conclusion, this study certifies that inflammatory factors in ITP patients significantly differ from the aged matched control group. The correlation significance of biomarkers with PLT were below the 0.01 level. However, this is a retrospective analysis and more and larger scale research is needed to confirm as the validity of such biomarkers.

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SUMMARY

PERIPHERAL BLOOD BIOMARKERS IN PATIENTS WITH REFRACTORY IMMUNE THROMBOCYTOPENIA

¹Metreveli S., ³Kvachadze I., ^{1,2}Kikodze N., ¹Chikovani T.,
¹Janikashvili N.

Tbilisi State Medical University, ¹Department of Immunology;
²Institute of Medical Biotechnology; ³Department of Physiology,
Georgia

Immune thrombocytopenia is an acquired thrombocytopenia caused by autoantibodies against platelet antigens, the pathogenesis of ITP is incompletely understood. Evidence regarding association between inflammatory factors in patients with ITP who are refractory to first line treatment is limited. The purpose of our study was to investigate the diagnostic value of NLR, PLR, and PMR, dNLR, and SII in ITP patients who were refractory for the first line treatment and had splenectomy as a second-line therapy.

Statistical analyses of inflammatory biomarkers were performed using SPSS v.26 and Graph Pad Prism. Correlations between the variables were determined by Spearman's correlation coefficient. The area under the curve (AUC), sensitivity, specificity, and cut-off values were compared using the receiver operating characteristic (ROC) curve.

Our data revealed that NLR and dNLR were increased ($p<0.0001$), while SII level was decreased ($p=0.0003$), PMR and PLR were also significantly low ($p<0.0001$) in ITP patients

compared with the age matched control group. In addition PLT level was negatively correlated with NLR and dNLR ($r=-0.605$, $P<0.01$), while there was positive correlation with SII, PLR, PMR (SII $r=0.799$; PLR $r=0.863$; PMR $r=0.40$, $P<0.01$). ROC curve analysis revealed AUC of PLR and PMR were 1.000 ($P=0.05$ and $P<0.001$), followed by SII 0.899 ($P=0.002$), NLR 0.875 ($P=0.04$) and dNLR 0.869 ($P=0.05$).

We observed that inflammatory factors in ITP patients significantly differ from the age matched controls, however larger scale research is needed to confirm the validity of such biomarkers.

Keywords: Immune thrombocytopenia, NLR, PLR, PMR, dNLR, SII, PLT.

РЕЗЮМЕ

БИОМАРКЕРЫ ПЕРИФЕРИЧЕСКОЙ КРОВИ У ПАЦИЕНТОВ С РЕФРАКТЕРНОЙ ИММУННОЙ ТРОМБОЦИТОПЕНИЕЙ

¹Метревели С.Д., ³Квачадзе И.Д., ^{1,2}Кикодзе Н.О.,

¹Чиковани Т.И., ¹Джаникашвили Н.Н.

*თბილისის სახელმწიფო მედიცინური უნივერსიტეტი,
/¹დეპარტამენტ იმუნოლოგია; ²ინსტიტუტ მედიცინურ ბიო-
ტექნოლოგია; ³დეპარტამენტ ფიზიოლოგია, საქართველო*

Иммунная тромбоцитопения (ITP) является приобретенной тромбоцитопенией, вызванной аутоантителами к антигенам тромбоцитов. Патогенез ITP до конца не изучен. Доказательства связи между воспалительными факторами у пациентов с ITP, не поддающихся лечению первой линии, ограничены.

Целью данного исследования явилась оценка диагностической ценности соотношений нейтрофилов и лимфоцитов (NLR), тромбоцитов и лимфоцитов (PLR), макрофагов и тромбоцитов (PMR), производного соотношения нейтрофилов и лимфоцитов (dNLR) и системного индекса иммунного воспаления (SII) у пациентов с ITP, рефрактерных к лечению первой линии и которым в качестве терапии второй линии была проведена спленэктомия.

Статистический анализ воспалительных биомаркеров проводился с использованием SPSS v.26 и Graph Pad Prism. Корреляции между переменными определялись коэффициентом корреляции Спирмена (Spearman). Исследованы площадь под кривой (AUC), чувствительность, специфичность и пороговое значение с использованием кривой рабочих характеристик приемника (ROC).

Анализ наших данных показал, что у пациентов с ITP, по сравнению с контрольной группой, показатели NLR и dNLR были статистически достоверно увеличены ($p<0.0001$), в то время как уровень SII был снижен ($p=0.0003$); PMR и PLR также были значительно низкими ($p<0.0001$). Кроме того, уровень PLT отрицательно коррелировал с NLR и dNLR ($r=-0.605$, $p<0.01$), в то же время отмечалась положительная корреляция с SII, PLR и PMR (SII $r=0.799$; PLR $r=0.863$; PMR $r=0.40$, $P<0.01$).

Анализ кривой ROC показал, что AUC для PLR и PMR составила 1.000 ($p=0.05$ и $p<0.001$), далее следовали SII - 0.899 ($P=0.002$), NLR - 0.875 ($P=0.04$) и NLR - 0.869 ($p=0.05$).

Таким образом, воспалительные факторы у пациентов с ITP значительно отличаются от показателей контрольной группы, однако для подтверждения достоверности данных биомаркеров необходимы более масштабные исследования.

რეზიუმე

პერიოდული სისხლის ბიომარკერები რეფრაქტორული იმუნური თრომბოციტოპენიის მქონე პაციენტებში

¹ს.ძეგლეველი, ²ო.გვაჭაძე, ^{1,2}ნ.ქიქოძე, ¹თ. ჩიქოვანი,
¹ნ.ჯანიგაშვილი

თბილისის სახელმწიფო სამედიცინო უნივერსიტეტი,
¹იმუნლოგიის დეპარტამენტი; ²სამდიცინო ბიოტექნიკლოგიის ინსტიტუტი; ³ფიზიოლოგიის დეპარტამენტი, საქართველო

იმუნური თრომბოციტოპენია (ITP) წარმოადგენს შექნილ თრომბოციტოპენიას რასაც თრომბოციტების საწინააღმდეგოდ მიმართული აუტოანტისტეულები იქვევს. ITP-ის პათოგენეზი ბოლომდე შესწავლილია არ არის. ლიტერატურაში მწირია მტკიცებულებანი სისტემური ანთებითი ფაქტორების როლის შესახებ პირველი რიგის მეურნალობის მიმართ რეფრაქტორული ITP-ს მქონე პაციენტებში. წარმოდგენილ კლიუშაში, სისტემური ანთების მარკერების საღიაგნოსტიკო მნიშვნელობის დადგნის მიზნით, რეფრაქტორული იმუნური თრომბოციტოპენიის მქონე პაციენტებში შეფასებულია პერიოდული სისხლიდან მიღებული ანთებადი ბიომარკერები: ნეიტროფილებისა და ლიმფოციტების ფარდობა (NLR), თრომბოციტებისა და ლიმფოციტების ფარდობა (PLR), თრომბოციტებისა და მონიციტების ფარდობა (PMR), ნეიტროფილებისა და ლიმფოციტების თანაფარდობის წარმოებული.

(dNLR) და სისტემური იმუნური ანთების ინდექსი (SII). სტატისტიკური ანალიზისთვის გამოყენებულია SPSS ვერსია 26 და Graph Pad Prism. ცვლადების შორის კორელაცია განისაზღვრა სპარმანის კორელაციის კოეფიციენტით. მიმდების ოპერატორული მასასიათურის (ROC) მრუდის საჭალებით განისაზღვრა მრუდის ქვედა ფარიობი (AUC), მგრძნობელობა, სპეციფიკურობა, ხდვული თპრიმადული მნიშვნელობა (cut-off Value). მიღებული შედეგების ანალიზში განჩვენა, რომ რეფრაქტორულ იტპ-კაციენტებში, საკონტროლო ჯგუფთან შედარებით, მნიშვნელოვნად მატულობს NLR და dNLR ($p < 0.001$), SII კი შემცირებულია ($p = 0.0003$), მნიშვნელოებანდ დაბალია ასევე PMR და PLR მაჩვენებლები ($p < 0.0001$); აღსანიშნავია, რომ თრომბოციტების რაოდენობა (PTL) უარყოფითად კორელირებს NLR და dNLR-მაჩვენებლებთან ($r = -0.605$, $p < 0.01$), ხოლო SII, PLR, PMR-თან მიმართებაში მისი კორელაცია დადგენითია (SII $r = 0.799$; PLR $r = 0.863$; PMR $r = 0.40$, $P < 0.01$). ROC მრუდის ანალიზში გამოავლინა, რომ PLR-ის და PMR-ის AUC-ს მაჩვენებელი მაქსიმალურია – 1.000 ($P = 0.05$ და $P < 0.001$), შემდგომ მოდის SII – 0.899 ($P = 0.002$), NLR – 0.875 ($P = 0.04$) და dNLR – 0.869 ($P = 0.05$). ამგარად, შეიძლება დაკასეკნათ, რომ რეფრაქტორული ITP-ს მქონე პაციენტებში სისტემური ანთებითი მარკერები მნიშვნელოვნად განსხვავდება შესაბამისი ასაკობრივი საკონტროლო ჯგუფის ანალიტიკური მაჩვენებლებისგან; თუმცა, ამ ბიომარკერების გაღიდობის დასაღასტურებლად უფრო მასშტაბური კვლევების ჩატარებაა აუცილებელი.

CLINICAL SONOGRAPHIC ANALYSIS OF BIOMETRIC INDICATORS OF BUCCAL THICKNESS AND BUCCAL FAT PAD IN PATIENTS WITH DIFFERENT FACIAL TYPES

¹Ruzhitska O., ²Kucher A., ³Vovk V., ¹Vovk Y., ³Pohranychna Kh.

Danylo Halych Lviv National Medical University, ¹Department of surgical and orthopedic dentistry of the faculty of postgraduate education; ²Department of Diagnostic Radiology of the Faculty of Postgraduate Education;
³Department of Surgical Dentistry and Maxillofacial Surgery, Ukraine

In recent years, the morphofunctional features of the buccal fat pad (BFP) of patients have been studied in detail for extensive clinical application in surgical dental practice. The morphological benefits of the BFP are related to its saturation with cell complexes with significant regenerative potential, high vascularization, malleable texture, plastic size preformation capabilities, and direct involvement in the implementation of the basic functions of the maxillofacial area (MFA). All this determines the potential for the application of the BFP to replace tissue defects and deformities [1-3]. Meanwhile, in literature and from our own experience with practical application of BFP, we encountered the issue of specific determination of its volumetric parameters in patients, which would significantly facilitate maxillofacial surgeons not only to remove a BFP fragment, but it would prevent the occurrence of unnecessary complications, such as postoperative hernia, pseudohernia, or BFP lipoma. From this perspective, we believe that, at the present stage of development of plastic and reconstructive surgery of the maxillofacial area, surgical approaches to the optimal and safe performance of such surgical interventions should be based on interdisciplinary assessment

of patients before surgery, and they take into account the individual typological characteristics of morphological facial features upon BFP sampling and grafting [4-6].

A clinical radiographic algorithm for the examination of patients with BFP sampling was clinically developed and statistically different parameters of the thickness of the buccal triangle were established in patients with different types of face [7,8]. X-ray examination with panoramic x-ray pictures revealed that the value of the sagittal angle and vertical distance from the occlusal surface of the first maxillary molar to the palatine plane also differed in patients with different types of face, which emphasizes the need for versatile clinical and instrumental improvement of preoperative diagnosis of BFP volumetric parameters.

Taking into account the foregoing, we consider that clinical and sonographic study of its biometric parameters in patients with different face typology is necessary for proper planning and efficient BFP sampling [9,10]. From this perspective, the purpose of our study was to establish a scientific and practical analysis of clinical sonographic results of the examination of patients with different types of face in preparation for reconstruction of the oral tissue defects of the BFP.